

This course will have four unique components. To successfully complete this course, students will need to submit pre-work assignments in a timely fashion and engage with asynchronous course materials. We will meet face to face the week of June 20-24, 2022 and will make efficient use of our time by completing laboratory activities, including analyses as well as other relevant activities.

The four components of the course are:

1. **Pre-work**–Asynchronous material will be posted to the course Canvas site at least one week prior to June 20. You may watch lectures and complete quizzes, worksheets, and other evaluation materials beginning June 13. Please note that some items will have deadlines associated with the in-person content.

2. **Evaluation** – Evaluation of your engagement and learning will include quizzes, worksheets, data analyses, and in person attendance and contributions to class. As noted above, some items will be bound to deadlines associated with our in-person modules.

3. **In-person** – Several hours per day have been designated for in-person attendance the week of June 20-24, 2022. Please review the schedule and plan to be on the Promega campus during that time.

4. **Office hours/Individual Consultations** – These can be scheduled as needed throughout the week of June 20-24, 2022

Date	Time	Module	Instructor				
Monday, June 20							
Asynchronous Pre-work and Evaluation Materials							
Please check Canvas for recordings which must be completed prior to attending in person and evaluation							
materials, which need to be completed prior to July 1.							
		Lecture	Sarah Teter				
		Purifying RNA					
		Lecture	Doug Horejsh				
		Studying miRNAs					
		Lecture	Rod Pennington				
		PCR Techniques with an					
		emphasis on RT-PCR and qPCR.					
Additional	To complete: Nucleic acid isolation quiz; Basic of Nucleic Acid Purification module;						
Assignments:	RNA Isolation worksheet; Real Time PCR quiz						
In-Person Schedule							
	9:00a – 9:30a	Welcome and Introductions	Amy Prevost, Erica Golueke				
	9:30a – 11:15a	Laboratory	Sarah Teter, Amy Prevost				
		RNA isolation from brain tissue					

Tentative Schedule:

		and spec analysis.	
	11:15a – 12:30p	Laboratory	Natalie Betz, Amy Prevost
		Cloning - RT-PCR amplification	
		Laboratory	
		RT-qPCR set up	
	12:30p – 1:00p	Lunch Lecture	Rod Pennington
		Questions about PCR?	
		Places where mistakes often	
		happen	
	1:00p – 3:00p	Laboratory	Erica Golueke, Michael Slater
		Transfect CRISPR pools for	
		studying protein degradation	
		Lecture	Michael Slater
		Using Cas-9 to create a double	
		strand break: Now what?!	
		CRISPR design basics.	
Tuesday. June 21			
Asynchronous Pre-	work and Evaluation	Materials	
Please check Canva	s for recordinas which	h must be completed prior to atte	ndina in person and evaluation
materials, which ne	ed to be completed p	rior to July 1.	
Additional	To complete: Clonir	ng quiz; Introduction to Gene Editi	ng module
Assignments:			-
In-Person Schedule	2		
	9:00a – 10:30a	Lecture	Jim Hartnett
		Cloning – Tools and	
		Techniques	
	10:30a – 11:15a	Laboratory	Natalie Betz, Amy Prevost
		Cloning – RT-PCR analysis and	
		ligation reaction	
		(Instructors will load and run	
		aels for students)	
	11·15a – 12·30n	lunchlecture	Michael Slater
	11.150 12.500	Consequences of double	Wiender Slater
		strand broaks, CDISDD	
		stration continued	
	12.20n 1.20n		Natalia Pota Amy Drayast
	12.20h – 1:20h	Cloping Transformation	watalle betz, Ally Prevost
	1.20n - 2.20n		Frica Colucko Amy Provest
	1.30p – 2.30p	Transfer CDICDD people to OC	Erica Golueke, Arity Prevost
		well plate	
		wen plate.	
	2:30p – 3:00p		Amy Prevost
		Cioning – plating cells	

Wednesday,				
June 22				
Asynchronous Pre-	work and Evaluation	Materials		
Please check Canva	s for recordings whicl	n must be completed prior to atter	nding in person and evaluation	
materials, which ne	ed to be completed p	rior to July 1.		
Additional	Complete Introduction to Bioluminescence Module			
Assignments:				
In-Person Schedule				
	9:00a – 10:30a	Lecture Changeland in Declaration	Mike Rosenblatt	
		Characterizing Proteins		
	10:30a – 11:30a	Lecture	Elizabeth Caine, Celia Bisbach	
	101000 111000	Studying protein degradation		
	11:30a – 12:30p	Lunch Lecture	Hicham Zegzouti	
		Kinase biology and drug		
		discovery		
	12:30p – 2:00p	Laboratory	Erica Golueke	
		Start kinetic read for PROTACs		
		Laboratory		
		Prepare cells for cell health		
	2.00 2.00-	analysis	TerryDise	
	2:00p-3:00p	Lecture Calls as reactions and	l erry Riss	
		Cells as reagents and		
Thursday, June 22				
Asynchronous Bro	work and Evaluation	Materials	L	
Asynchronous Pre-	s for recordings which	n must be completed prior to atte	nding in person and evaluation	
materials. which ne	ed to be completed p	rior to July 1.	nang in person ana evaluation	
.,		Lecture	Chris Eggers	
		Western Blot, ICC and ELISA		
Additional			•	
Assignments:				
In-Person Schedule	2			
	9:00a – 10:00a	Lecture	Chris Eggers	
		Basics of Western Blotting		
	10:00a – 11:30a	Laboratory	Chad Zimprich, Amy Prevost	
		Western Blot: prepare and run		
		gels; begin transfer		

	11:30a – 12:15p	Laboratory	Natalie Betz, Amy Prevost
		Analysis of RT-qPCR Data	
		Western Blot: Blocking and	Chad Zimprich, Amy Prevost
		primary antibody addition	
	12:15p – 1:30p	Lunch Discussion	Elizabeth Caine. Celia
		Review basics of studying	Bisbach, Michael Slater
		protein degradation + PROTAC	
		data analysis & remaining	
		guestion about genome	
		editing	
	1:30p – 3:00p	Laboratory	Chad Zimprich, Amy Prevost
		Western Blot: Washes (15 min)	
		& apply secondary antibody	
		(30 min. during which we will	
		switch to)	
		,	
		Laboratory: Start Colony PCR	Natalie Betz. Amy Prevost
		,,	
		Laboratory	Chad Zimprich, Amy Prevost
		Western Blot: Wash and	, , , ,
		develop	
Friday, June 24			
Asynchronous Pre-	work and Evaluation	Materials	
Please check Canvas	s for recordinas which	n must be completed prior to atter	nding in person and evaluation
materials, which ne	ed to be completed p	rior to July 1.	5 /
Additional	Complete Cell Healt	th Quiz; complete delta-delta Cg a	nalysis worksheet
Assignments:			,
In-Person Schedule			
	9:00a – 10:30a	Lecture	Andrew Niles
		Studving cell death and	
		monitoring cell health: Using	
		NanoLuc to track apoptotic	
		events in cells	
	10:30a – 1:30n	Laboratory	Frica Golueke, Andrew Niles
		Monitoring cell health	
		(Pizza lunch)	
		Laboratory	Natalie Betz, Amy Prevost
		Gel analysis of colony PCR	
	1:30p-2:30p	Guest lecture	Reid Alisch
	1.000 2.000		
		Foigenetics and	
		Epigenetics and Neurodevelopment	
	2.30n - 2.00n	Neurodevelopment	